Page 10, third full paragraph, which wraps to page 11:

In a first set of optimization experiments, efficient incorporation of pal-prot A was documented in four cell lines (Fig. 1*A*) as detected with FITC-conjugated human IgG. As a negative control, nonderivatized protein A lacked the capacity to bind to the same cells. Data from the FACStar® flow cytometer analysis was plotted as arbitrary units of log10 fluorescence intensity versus number of EL-4 cells; membrane incorporation was dose dependent and started to plateau at about 33 μg/ml pal-prot A, as shown in Fig. 1*B*. EL-4 cells were incubated with 33 μg/ml pal-prot A for the indicated periods of time and processed as above; pal-prot A incorporation was rapid, appearing immediately after addition to the cells and reaching a plateau at ~1 h, as shown in Fig. 1*C*. This data demonstrates that numerous different cell lines can be used in the present protein transfer methods, and that the lipidated protein was incorporated into the cell fairly rapidly.

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Page 11, last paragraph:

Cells precoated with pal-prot A were washed once and resuspended in RPMI 1640 medium (3-7 x 1^6 cells/ml). pREP7B-transfected K562 cells (K562/REP7b) were serially incubated with 33 μg/ml protein A for 2 h, 33 μg/ml $Fc\gamma_1$ fusion protein for 1 h, and BB-1 as primary Ab and FITC-conjugated goat anti-mouse IgG as secondary Ab. To monitor protein delivery, 10^6 cells were washed twice with the same buffer as above, incubated on ice for 1 h with 1 μg of human B7-specific mAb BB-1 (PharMingen, San Diego, CA) in 100 μl of buffer. Cells were washed once and immunostained (on ice for 1 h) with 100 μl of 1:100 diluted FITC-conjugated goat $F(ab')_2$ anti-mouse Ig (Boehringer Mannheim, Indianapolis, IN) as secondary Ab. Cells were washed once, resuspended in PBS, and analyzed on a FACStar® flow cytometer.

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Page 13, second full paragraph

PBMC were isolated from fresh whole blood by Ficoll density centrifugation. T-cells were purified by two rounds of treatment with Lympho-kwik (One Lambda, Canoga Park, CA). T-cell purity was verified by lack of a proliferative response to phytohemaglutin ("PHA") or PMA in the absence of accessory cells. The human CD3-specific mAb HIT3a (PharMingen) was bound to 96-well plates at the indicated concentrations and used in this form to provide a first activating signal to T-cells. Alternatively, PHA was used in soluble form as a source of a first signal. K562 cells transfected with the negative control vector pREP7β (K562/pREP7β) were precoated with

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